NOTES

Quinocitrinines A and B, New Quinoline Alkaloids from *Penicillium citrinum* Thom 1910, a Permafrost Fungus

A. G. Kozlovsky, V. P. Zhelifonova, T. V. Antipova, V. M. Adanin, S. M. Ozerskaya and G. A. Kochkina

G.K. Skrjabin-Institute of Biochemistry and Physiology of Microorganisms, Russian Academy of Sciences, 142290 Pushchino, Moscow Region, Russia

B. SCHLEGEL, H. M. DAHSE, F. A. GOLLMICK and U. GRÄFE

Hans-Knöll-Institute for Natural Products Research, Beutenbergstrasse 11, D-07745 Jena, Germany

(Received for publication February 4, 2003)

In the course of our previous work we isolated alkaloids such as agroclavine-I and epoxy-agroclavine-I from *Penicillium citrinum* VKM FW-800 as a microorganism isolated from permafrost^{1,2)}. This fungus was shown to coproduce two other alkaloid compounds, which were extractable only under acidic conditions. Positive staining by Dragendorff's reagent but negative Ehrlich reaction suggested the occurrence of unusual alkaloid structures. Here we report isolation and structure elucidation of quinocitrinines A (1) and B (2) as new quinoline alkaloids from *Penicillium citrinum*.

Penicillium citrinum VKM FW-800 obtained from the permafrost region of Northern Russia was deposited in the All Russian Culture Collection of Microorganisms (VKM, Pushchino, Moscow region, Russia). Cultivation was carried out at 24°C on rotary shaker (220 r.p.m.) in Erlenmeyer flasks (750 ml) containing 150 ml medium. The medium was composed as follows (g/liter): mannitol 50, succinic acid 5.4, MgSO₄·7H₂O 0.3, KH₂PO₄ 1.0 (pH adjusted to 5.2 by addition of 25% ammonia solution). Inoculation occurred with spore suspensions. After 11~13 days of cultivation the metabolites were extracted. The culture broth was adjusted to pH 3~4 by 2% tartaric acid and extracted three-times by chloroform. The residue of the

dried and evaporated extract was subjected to column chromatography on silica gel 60 (Merck, $0.063 \sim 0.1$ mm). Elution was done first by CHCl₃/MeOH/25% aqueous ammonia (90:10:0.1; v/v) and subsequently by the same solvent mixture but in ratio 80:20:0.2 (v/v).

Thereby 15.2 mg of **1** and 40.1 mg of **2** were obtained as slightly brownish solids. The physico-chemical properties of **1** and **2** are shown in Table 1.

Elucidation of structure of **1** and **2** (Fig. 1) was carried out by optical spectroscopy (UV-VIS, IR, polarimetry), mass spectrometry (HREI-MS, ESI-MS) and 1D/2D NMR spectroscopy.

Quinocitrinines A (1) and B (2) differed by their chromatographic behaviour (Rf on TLC), melting points and optical rotations. However the UV-Vis spectra were identical but distinguishable from those of typical ergot alkaloids. Absorbances at $300 \sim 330$ nm suggested the occurrence of a quinoline chromophore. 1 and 2 displayed different optical rotations (Table 1). The identical IR spectra of both alkaloids displayed absorbances (cm⁻¹) typical for carbonyl (1690 cm⁻¹), hydroxyl (3245 cm⁻¹) and double bonds (1605 cm⁻¹).

ESI-MS of 1 afforded m/z 293.3 ([M+Na]⁺) and of 2 m/z 293.5 ([M+Na]⁺). Additionally in both spectra m/z 563 ([2M+Na]⁺) was visible. HREI-MS of 1 furnished m/z 270.1373 ([M-H]⁺; calcd. 270.1379 for C₁₆H₁₈N₂O₂), and of 2 m/z 270.1374 ([M-H]⁺; calcd. 270.1379 for C₁₆H₁₈N₂O₂). The appearance of [M-H]⁺ ions in the HREI-MS of both compounds is in full accord with the

Fig. 1. Structure (relative stereochemistry tentative) of quinocitrinines A (1) and B (2).



^{*} Corresponding author: gollmick@pmail.hki-jena.de

| | 1 | 2 |
|--|-------------------------|--------------------|
| Appearance: | slightly brownish solid | slightly brownish |
| | solid | solid |
| Mp (°C): | 142-143 | 152-153 |
| Molecular weight: | 270 | 270 |
| HREI-MS found: | 270.1373 | 270.1374 |
| calcd.: | 270.1379 | 270.1379 |
| Molecular formula: | $C_{16}H_{18}N_2O_2$ | $C_{16}H_8N_2O_2$ |
| UV-VIS MeOH, $\lambda_{max}(nm)$: | | |
| ϵ (cm ² /mmol) | 216(8108), 248(5285), | 216 (8108), 248 |
| | 256(5430), 300 (2388), | (5772), 256 (5831) |
| | 300(2388), 314(2785), | 300 (2542), 314 |
| | 328(2499) | (3074), 328 (1973) |
| IR film, v_{max} (cm ⁻¹): | 3263, 1689, 1604, 1545 | , 3245, 1690,1605, |
| | 1523, 1462, 1420 | 1545, 1524, 1462, |
| | | 1420 |
| $[\alpha]_{D}^{22}$ (c 0.32, MeOH): | +25.3° | |
| $[\alpha]_{D}^{22}$ (c 0.62, MeOH): | | -13.2° |
| R _f on TLC (silical gel alumini | um | |
| sheets, Merck) | | |
| Solvent 1: CHCl ₃ -MeOH-cond | 0.68 | 0.62 |
| NH ₄ OH: (80:20:0.2): | | |
| Solvent 2: CHCl ₃ -MeOH-cond | o. 0.49 | 0.45 |
| NH ₄ OH (90:10:0.1): | | |
| Solvent 3: CHCl ₃ -MeOH-cond | 0.14 | 0.12 |
| NH4OH (90:10:1) | | |

.

| Table 1. Physico-chemical properties of quinocitinine A (1) and quinocitinine B (| Fable | 1. | Physico-chemical | properties of c | juinocitrinine A (| 1) and c | quinocitrinine B | (2). |
|---|-------|----|------------------|-----------------|--------------------|----------|------------------|------|
|---|-------|----|------------------|-----------------|--------------------|----------|------------------|------|

occurrence of quaternary ammonium structures. The chemical formulas of **1** and **2** suggested the presence of nine double bonds or rings in the molecules.

Conclusive evidence for the structure of **2** as shown in Fig. 1 was furnished by 1D and 2D NMR spectroscopy (¹H, ¹³C, DEPT, HMQC, HMBC, NOESY, in pyridine- d_5). However using CDCl₃ as solvent several carbons and C,H long-range couplings in the spectra of both compounds were invisible. The NMR spectra of **1** and **2**, too, were fully identical supporting the view that both compounds are diastereomers. In the ¹³C and DEPT NMR spectra (Table 2) 16 carbon atoms and their bonding types were visible (five CH or CH₃ groups, one methylene structure (21.3 ppm), 4 doubly bonded C–H groups, and six quaternary double

bond carbons). Downfield shift of three of the latter carbons (C-9: 169.9 ppm, C-3: 172.5 ppm, C-10: 164.7 ppm) proposed the presence of carbon-heteroatom bonds. If the ¹³C and 2D NMR spectra of **1** and **2** were recorded in chloroform the aromatic carbon signals of C-6, C-7 and C-8 were not seen in accord with the occurrence of tautomeric forms constituting the molecules under ambient conditions. In the same manner the unusual downfield shift of C-3 (172.5 ppm) was explainable.

The ¹H NMR and ¹H, ¹H-COSY NMR spectra showed the presence of two *ortho*-coupled (H-4, H-5) and two *meta*-coupled aromatic protons (H-2, H-7). The aliphatic side chain at C-11 was fully assignable due to the observable shift data and coupling pattern. Additional proton signals at

| position | δ _C | δ _H |
|----------|----------------|---|
| 1 | 141.7(s) | |
| 2 | 116.7 (d) | 7.60 d, 1.5 |
| 3 | 172.5 (s) | |
| 4 | 124.8 (d) | 7.38 dd, 1.5; 7.4 |
| 5 | 126.8 (d) | 8.70 dd, 7.4, 1.0 |
| 6 | 128.9 | |
| 7 | 132.7 (d) | 7.60 d, 1.0 |
| 8 | 111.0 (s) | <u> </u> |
| 9 | 169.9 (s) | |
| 10 | 164.7 (s) | _ |
| 11 | 60.2 (d) | 4.88 m |
| 12 | 36.7 (d) | 2.21 m |
| 13 | 21.3 (t) | $H_{\rm A}$ 0,.82 m; $H_{\rm B}$ 0.90 m |
| 14 | 11.5 (q) | 0.49 t, 7.1 |
| 15 | 17.8 (q) | 1.21 d, 6.9 |
| 16 | 36.8 (q) | 3.60 s |

Table 2. Assignment of the identical ¹H and ¹³C NMR spectra quinocitrinines A (1) and B.

In pyridine- d_5 , chemical shifts (δ) in ppm; abbreviations: s: singlet, d: doublet, t: triplet, q: quartet, m: multiplet, br: broad, coupling constants in Hz.

Fig. 2. Instructive C,H long-range couplings (HMBC, arrows at one side) and NOESY correlations (arrows at both sides) in the NMR spectra of 1.



9.20 ppm (N–H, s; broad) and 3.60 ppm (s, 3H) were attributable to a nitrogen-bonded lactam proton and, respectively at methyl group (C-16) attached to the quaternary ring nitrogen.

For full structural assignment the 2D spectra such as HMQC and HMBC were of pivotal importance. C,H longrange couplings in the HMBC spectra of 1 and 2 (Fig. 2) confirmed doubtlessly the structures of 1 and 2. Thus diagnostic couplings of H-16 with C-1 and C-11, of H-5 with C-7, of H-11 with C-8 and C-10, and of the nitrogenbonded proton at 9.20 ppm with C-8 and C-11 (Fig. 2) supplied supporting evidence. Due to the identity of UV-VIS, IR and NMR spectra of 1 and 2, and the differences in Rf values, optical rotation and melting point (Table 1) it can be suggested that both compounds possess diastereomeric structures at the two stereocenters (C-11, C-12). However the differences in optical rotation values will not enable the assignment of absolute configuration for one of the stereocenters. The relative stereochemistry (see Fig. 1) was based on the assumption that D- or L-isoleucine forms the precursors for the aliphatic part of the molecules. The quinoline ring and the attached carbonyl group (C-9) can be suggested to originate from tyrosine.

1 and 2 display moderate antibacterial activities against a spectrum of Gram-positive and Gram-negative bacteria, and fungi (Table 3). Moreover, moderate antiproliferative

| Test organism | IZ (1 ; mm) | IZ (2 ; mm) |
|-------------------------|-------------|---------------------|
| Bacillus subtilis ATCC | 13 | 12 |
| 6633 | | |
| Staphylococcus aureus | 12 | 13 |
| SG511 | | |
| Escherichia coli SG 458 | 14 | 14 |
| Pseudomonas aeruginosa | 15 | 15* |
| SG137 | | |
| Pseudomonas aeruginosa | 0 | 0 |
| K599/61 | | |
| Staphylococcus aureus | 11 | 11* |
| 134/9 | | |
| Enterococcus faecalis | 12 | 12 |
| 1528 | | |
| Mycobacterium | 13 | 12* |
| smegmatis SG987 | | |
| Sporobolomyces | 0 | 0 |
| almonicolor 549 | | |
| Candida albicans Bay R. | 12 | 12 |
| Penicillium sp. JP 36 | 15 | 15* |

Table 3. Antimicrobial activities of 1 and 2.

 $50 \,\mu$ l of 1 mg/ml solutions were added to agar wells, and the diameter of inhibition zones (IZ) was determined in mm (* means diffuse inhibition halo).

activity (IC₅₀; μ g/ml) was found with L-929 cells (1: 33.1; 2: 18.6), K-562-cells (1: 19.5; 2: 7.8) and HeLa cells (1: >50; 2: >50).

The physico-chemical data thus show quinocitrinines A (1) and B (2) as new bioactive representatives of the rare quinoline-type of fungal alkaloides.

Experimental

UV-VIS spectra were recorded on a Specord 2000 double beam instrument (Analytik Jena, Germany), FTIR spectra on a Mattson Satellite FTIR spectrometer equipped with ATR device (Mattson, Chicago, USA), optical rotation on a Propolis polarimeter (Dr. KERNCHEN OPTICS, Seelze, Germany), ESI mass spectra on a triple quadrupole mass spectrometer Quattro (VG Biotech, Altrincham England), EREI mass spectra on a Finnigan MAT 95XL (Finnigan Bremen, Germany) and NMR spectra on a Bruker Avance DRX 500 (Bruker, Karlsruhe, Germany).

Acknowledgements

Support of this work by Deutsche Forschungsgemeinschaft Bonn (436 RUS 113/677/1-1) is gratefully acknowledged.

References

- KOZLOVSKY, A. G.; T. F. SOLOVIYOVA, V. G. SAKHAROVSKY & V. M. ADANIN: Ergot alkaloids agroclavine-I and epoxyagroclavine-I—metabolites of *Penicillium corilophillum*. Prikladnaya biokhimia i mikrobiologiya. VXVIII: 4, 535~541, 1982, (in Russian)
- 2) KOZLOVSKY, A. G.; V. P. ZHELIFONOVA, V. M. ADANIN, T. V. ANTIPOVA, S. M. OZERSKAYA, G. A. KOCHKINA & U. GRÄFE: Filamentous fungus Thom 1910 VKM FW-800, isolated from ancient permafrost as a producer of agroclavine-I and epoxyagroclavine-I. Mikrobiologiya, (in Russian, in press) 2002